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Biomimetic Synthesis Very Important Paper

Biomimetic Total Synthesis of Hyperjapones A-E and Hyperjaponols A and C

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Abstract: Hyperjapones A–E and hyperjaponols A–C are complex natural products of mixed aromatic polyketide and terpene biosynthetic origin that have recently been isolated from Hypericum japonicum. We have synthesized hyperjapones A-E using a biomimetic, oxidative hetero-Diels-Alder reaction to couple together dearomatized acylphloroglucinol and cyclic terpene natural products. Hyperjapone A is proposed to be the biosynthetic precursor of hyperjaponol C through a sequence of: 1) epoxidation; 2) acid-catalyzed epoxide ring-opening; and 3) a concerted, asynchronous alkene cyclization and 1,2-alkyl shift of a tertiary carbocation. Chemical mimicry of this proposed biosynthetic sequence allowed a concise total synthesis of hyperjaponol C to be completed in which six carbon-carbon bonds, six stereocenters, and three rings were constructed in just four steps.

Typerjapones A- $E^{[1]}$ (Figure 1, 1-5) are a family of five structurally related meroterpenoids isolated from Hypericum japonicum, a plant used to treat hepatitis in traditional Chinese medicine. Hyperjapone A is a racemic natural product with an 11-6-6 tricyclic ring system. Hyperjapones B-E are enantiopure natural products which share

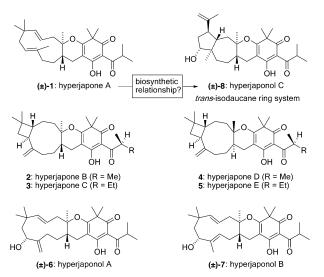


Figure 1. Hyperjapones A-E and hyperjaponols A-C.

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a common 4-9-6-6 ring system. Further racemic natural products, hyperjaponols A-C^[2] (6-8), have also been isolated from Hypericum japonicum. The most structurally intriguing of these natural products is hyperjaponol C (8) because its stereochemically complex, but racemic, trans-isodaucane framework implies a highly predisposed, non-enzymatic biosynthesis.

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Our detailed proposal for the biosynthesis of hyperjapone A (1) and its conversion into hyperjaponols A-C (6-8) is presented in Scheme 1. First, trimethylation of acylphloroglucinol 9 would give the dearomatized natural product norflavesone (10).^[3] Oxidation of 10 would give the α,β unsaturated ketone 12, which could be expected to exhibit similar reactivity to an o-quinone methide.[4] It could therefore undergo a non-enzymatic, hetero-Diels-Alder reaction with humulene (11) to give racemic hyperjapone A (1). The $\Delta^{1,2}$ alkene of humulene was shown to be the most reactive dieneophile in hetero-Diels-Alder reactions between humulene and an o-quinone methide in both the biomimetic synthesis of lucidene by Baldwin et al. [5] and the biomimetic synthesis of guajadial B by Liu and co-workers. [6] The biosynthesis of hyperjapone A (1) should therefore be inherently regioselective with respect to humulene (11). The hetero-Diels-Alder reaction would also be expected to be regioselective with respect to the α,β -unsaturated ketone 12 as its most stable tautomer would be stabilized by an intramolecular hydrogen bond (as shown in Scheme 1). Hyperjapones B-E (2-5) could arise from a similar nonenzymatic, oxidative hetero-Diels-Alder reaction between a trimethylated acylphloroglucinol and the reactive trans $\Delta^{4,5}$ alkene of carvophyllene (18).[7]

The X-ray crystal structure of hyperjapone A (1) shows that it adopts a conformation with one face of the $\Delta^{8,9}$ alkene exposed, while the other face is sterically hindered inside the 11-membered ring (Scheme 1).^[1] Diastereoselective epoxidation of the exposed face of the $\Delta^{8,9}$ alkene would therefore give epoxide 13, which has a 1R*2R*8S*9S* relative configuration. A similarly regio- and diastereoselective result was obtained in a recent diepoxidation of humulene (11) by Fujita and co-workers, in which the structure of the diepoxide product was elucidated using the crystalline sponge method.^[8] Acid-catalyzed ring opening of epoxide 13 would generate tertiary carbocation 14, with the carbocation centered at the C9 position. Deprotonation of 14 could then give either hyperjaponol A (6) or hyperjaponol B (7). Alternatively, a stereoselective cation-alkene cyclization of 14 could give secondary carbocation 15, with the relative configuration of the newly formed trans ring junction dictated by the orientation of the $\Delta^{4,5}$ alkene in the favored hyperjapone A conformation. A stereospecific 1,2-alkyl shift of secondary





Scheme 1. Proposed biosynthesis of hyperjaponol C from hyperjapone A through a concerted, asynchronous alkene cyclization/1,2-alkyl shift.

carbocation **15** would then generate tertiary carbocation **16**, and a final deprotonation would give hyperjaponol C (8). Alternatively, it is perhaps more likely that carbocation **14** would directly rearrange to give carbocation **16** through a concerted, asynchronous alkene cyclization/1,2-alkyl shift. Similar concerted carbocation rearrangements in terpene biosynthesis have been proposed and studied computationally by Tantillo and co-workers.^[9]

The primary aim of this project was to use the biosynthetic hypothesis outlined in Scheme 1 as the blueprint for a divergent, biomimetic synthesis of hyperjaponols A-C (6-8) from hyperjapone A (1). Our synthesis of hyperjapone A began with Friedel-Crafts acylation of phloroglucinol (17) with isobutyryl chloride to give an acylphloroglucinol, [10] which was dearomatized by means of trimethylation^[11] to give norflavesone (10) in good yield over two steps (Scheme 2). Oxidation of 10 with TEMPO and Ag₂O in the presence of humulene (11) then generated hyperjapone A (1) in 32% yield, presumably through a hetero-Diels-Alder reaction between the α,β -unsaturated ketone 12 generated in situ and humulene (11). The use of TEMPO and Ag₂O in tandem was the result of extensive reaction optimization (see the Supporting Information for full details). Ag₂O has previously been used for the oxidative generation of o-quinone methides, $^{[12]}$ but not in the oxidative formation of α,β -unsaturated ketones. We have previously used a combination of TEMPO and PhI- $(OAc)_2$ to induce a similar oxidation to generate an α,β unsaturated ketone that underwent a 6π -electrocyclization in our biomimetic synthesis of hyperguinone B.^[13]

With a concise synthesis of hyperjapone A (1) established, we next focused on its biomimetic conversion into hyper-

Scheme 2. Total synthesis of hyperjapone A and hyperjaponols A and C. TEMPO = (2,2,6,6-tetramethylpiperidin-1-yl)oxyl; m-CPBA = 3-chloroperoxy-benzoic acid; p-TsOH = p-toluenesulfonic acid.

japonol C (8). As expected from molecular modelling studies and Fujita's humulene epoxidation work, [8] treatment of 1 with m-CPBA generated epoxide 13 as a single diastereomer in 76% yield through diastereoselective oxidation of the $\Delta^{8,9}$ alkene. We were then delighted to observe that acid-





catalyzed rearrangement of 13 using p-TsOH in CH₂Cl₂ gave hyperjaponol C (8) in 43% yield, presumably by the concerted, asynchronous alkene cyclization/1,2-alkyl shift mechanism outlined in Scheme 1. The use of AgNO₃/SiO₂ chromatography was essential in the purification of 8 from trace reaction by-products.^[14] The synthesis of **8** could be streamlined by combining the epoxidation and acid-catalyzed rearrangement steps into a one-pot procedure, which directly converts 1 into 8 in 34 % yield. [15] This operationally simple reaction diastereoselectively generates four stereocenters of the unusual trans-isodaucane skeleton of 8. Almost all isodaucane terpenoid natural products previously isolated possess a cis-fused ring junction that is likely to be biosynthesized through cyclization of a germacrene D derivative. [16] Several acid-catalyzed rearrangements of humulene^[17] and humulene-8,9-epoxide^[18] that are similar to the conversion of 1 into 8 have been previously reported. However, these rearrangements are generally unselective, giving rise to complex mixtures of products. Furthermore, our synthesis of 8 is the first time that this rearrangement mode of a humulene derivative has been used in a natural product synthesis, and it is the first time that it has been proposed to occur in a biosynthetic pathway.

Conversion of epoxide 13 into hyperjaponol A (6) was achieved in 59% yield upon treatment with (NC)₂C=C(CN)₂ and LiBr in acetone. [19] Despite extensive screening of further Lewis and protic acids, the formation of hyperjaponol B (7) from 13 has not yet been observed.

The secondary aim of this project was to synthesize hyperjapones B-E using an oxidative hetero-Diels-Alder reaction to couple caryophyllene (18) with trimethylated acylphloroglucinol natural products. Thus, treatment of norflavesone (10) with TEMPO/Ag2O in the presence of caryophyllene (18) gave a 2.5:1 mixture of hyperjapone B (2) and hyperjapone D (4) in 60% combined yield (Scheme 3). 18 exists as a 3:1 mixture of $\beta\alpha$ and $\beta\beta$

Scheme 3. Total synthesis of hyperjapones B and D.

conformations in solution. [20] 2 is formed by the cycloaddition of the α,β -unsaturated ketone 12 generated in situ to the reactive \textit{trans} \, \Delta^{4,5} alkene of the more abundant $\beta\alpha$ conformation, while 4 is generated from addition to the less abundant ββ conformation.^[7] In their isolation paper, Xu et al. reported the separation of 2 and 4 using preparative HPLC.^[1] However, we also found that 2 could be purified from 4 by means of selective crystallization from MeOH.

Scheme 4. Total synthesis of hyperjapones C and E.

The total syntheses of hyperjapones C (3) and E (5) was achieved in a similar fashion (Scheme 4). Friedel-Crafts acylation of phloroglucinol (17) with (S)-2-methylbutyryl chloride^[21] followed by trimethylation gave the dearomatized natural product norisoleptospermone (19). Exposure of 19 to TEMPO/Ag₂O in the presence of caryophyllene (18) gave a 2.5:1 mixture of hyperjapone C (3) and hyperjapone E (5) in 61% combined yield. Again, the product ratio of the oxidative hetero-Diels-Alder reaction approximately correlates to the ratio of the $\beta\alpha$ and $\beta\beta$ conformations of 18.

Additionally, we synthesized 20 and 21 as an inseparable 1:1 mixture of diastereomers by treatment of 19 with TEMPO/Ag₂O in the presence of humulene (11). Given the probable biosynthesis of hyperjapones A-E in Hypericum japoniucm through non-enzymatic hetero-Diels-Alder reactions, it is highly likely that 20 and 21 are "undiscovered natural products". Samples of 20 and 21, and also hyperjapone A epoxide (13), have therefore been distributed to isolation chemists in order to accelerate their discovery in nature.

In conclusion, we have used a biomimetic oxidative hetero-Diels-Alder reaction to synthesize hyperjapones A-E (1-5). This strategy allows for efficient coupling of dearomatized, trimethylated acylphloroglucinol natural products to the reactive trans alkenes of cyclic terpenes. Hyperjapone A (1) was converted into hyperjaponol A (6) and hyperjaponol C (8) through acid-catalyzed rearrangement of an intermediate epoxide. The four-step synthesis of hyperjaponol C (8) from simple starting materials involves the construction of six carbon-carbon bonds, six stereocenters, and three rings, and is thus a good example of the use of a biomimetic synthetic approach to rapidly generate molecular complexity. More generally, this work shows that a biomimetic approach to synthesis can lead to both naturally divergent strategies (nine natural products and three proposed "undiscovered natural products" have been synthesized, all for the first time) and naturally efficient syntheses

Communications





(all routes are protecting-group-free, with excellent step, redox, and pot economy).

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- [1] X.-W. Yang, Y.-P. Li, J. Su., W.-G. Ma, G. Xu, Org. Lett. 2016, 18, 1876
- [2] L. Hu, Y. Zhang, H. Zhu, J. Liu, H. Li, X.-N. Li, W. Sun, J. Zeng, Y. Xue, Y. Zhang, Org. Lett. 2016, 18, 2272.
- [3] D. P. Killeen, L. Larsen, F. E. Dayan, K. C. Gordon, N. B. Perry, J. W. van Klink, J. Nat. Prod. 2016, 79, 564.
- [4] For reviews of o-quinone methide chemistry, see: a) R. W. Van De Water, T. R. R. Pettus, Tetrahedron 2002, 58, 5367;
 b) N. J. Willis, C. D. Bray, Chem. Eur. J. 2012, 18, 9160;
 c) W.-J. Bai, J. G. David, Z.-G. Feng, M. G. Weaver, K.-L. Wu, T. R. R. Pettus, Acc. Chem. Res. 2014, 47, 3655.
- [5] a) R. M. Adlington, J. E. Baldwin, G. J. Pritchard, A. J. Williams, D. J. Watkin, *Org. Lett.* 1999, 1, 1937; b) R. Rodriguez, J. E. Moses, R. M. Adlington, J. E. Baldwin, *Org. Biomol. Chem.* 2005, 3, 3488.
- [6] Y. Gao, G.-Q. Wang, K. Wei, P. Hai, F. Wang, J.-K. Liu, Org. Lett. 2012, 14, 5936.
- [7] For examples of biomimetic hetero-Diels-Alder reactions involving caryophyllene, see: a) A. L. Lawrence, R. M. Adlington, J. E. Baldwin, V. Lee, J. A. Kershaw, A. L. Thompson, *Org. Lett.* 2010, 12, 1676; b) J. T. J. Spence, J. H. George, *Org. Lett.*

- **2011**, *13*, 5318; c) K. Takao, S. Noguchi, S. Sakamoto, M. Kimura, K. Yoshida, K. Tadano, *J. Am. Chem. Soc.* **2015**, *137*, 15971.
- [8] N. Zigon, M. Hoshino, S. Yoshioka, Y. Inokuma, M. Fujita, Angew. Chem. Int. Ed. 2015, 54, 9033; Angew. Chem. 2015, 127, 9161.
- [9] a) D. J. Tantillo, Chem. Soc. Rev. 2010, 39, 2847; b) D. J. Tantillo, Nat. Prod. Rev. 2011, 28, 1035.
- [10] L. Crombie, R. C. F. Jones, C. Palmer, J. Chem. Soc. Perkin Trans. 1 1987, 317.
- [11] N. T. Nguyen, V. C. Pham, M. Litaudon, F. Guéritte, B. Bodo, V. T. Nguyen, V. H. Nguyen, *Tetrahedron* 2009, 65, 7171.
- [12] D. Liao, H. Li, X. Lei, Org. Lett. 2012, 14, 18.
- [13] J. H. George, M. D. Hesse, J. E. Baldwin, R. M. Adlington, *Org. Lett.* 2010, 12, 3532.
- [14] a) C. M. Williams, L. N. Mander, *Tetrahedron* 2001, *57*, 425;
 b) L. N. Mander, C. M. Williams, *Tetrahedron* 2016, *72*, 1133.
- [15] For a similarly telescoped one-pot reaction involving epoxidation followed by acid-catalyzed rearrangement, see: A. W. Markwell-Heys, K. K. W. Kuan, J. H. George, *Org. Lett.* 2015, 17, 4228.
- [16] N. Bülow, W. A. König, Phytochemistry 2000, 55, 141.
- [17] a) Y. Naya, Y. Hirose, Chem. Lett. 1973, 2, 133; b) Y. Naya, Y. Hirose, Chem. Lett. 1973, 2, 727; c) D. Baines, J. Forrester, W. Parker, J. Chem. Soc. Perkin Trans. 1 1974, 1598; d) W. G. Dauben, J. P. Hubbell, N. D. Vietmeyer, J. Org. Chem. 1975, 40, 479.
- [18] I. Bryson, J. S. Roberts, A. Sattar, Tetrahedron Lett. 1980, 21, 201.
- [19] I. G. Collado, J. R. Hanson, A. J. Macías-Sánchez, *Tetrahedron* 1996, 52, 7961.
- [20] M. Clericuzio, G. Alagona, C. Ghio, L. Toma, J. Org. Chem. 2000, 65, 6910.
- [21] M. J. Begley, L. Crombie, R. C. F. Jones, C. J. Palmer, J. Chem. Soc. Perkin Trans. 1 1987, 353.

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